

## STEROLS FROM THE LEAFY CULMS OF *Desmostachya bipinnata*

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*Desmostachya bipinnata* (L.) Stapf (Poaceae), also known as sacrificial grass, is a perennial herbaceous plant distributed from north Africa to South Asia [1]. The leafy culms and roots of the plant are used as an aphrodisiac and diuretic in traditional Ayurvedic medicine [2]. *D. bipinnata* has also been reported to have antifungal [3] and antiurolithiatic [4] activities. So far, flavonoids such as kaempferol, quercetin, quercetin-3-glucoside, trycin, trycin-7-glucoside, and 4'-methoxy quercetin-7-O-glucoside have been reported from this plant [5, 6]. *D. bipinnata* has pharmacological components such as trycin and trycin-7-glucoside, which are reported to be anti-ulcerogenic [5], and 4'-methoxy quercetin-7-O-glucoside, which is reported as an anti-helicobacter [6].

Fresh leafy culms of *D. bipinnata* were collected from Birgunj, Nepal in August 2008. The dried leafy culms of *D. bipinnata* (1.3 kg) were extracted with an 80% aqueous MeOH (20 L × 3) solution at room temperature. The concentrated extracts were successively partitioned with water (3.5 L), EtOAc (3 L × 3), and *n*-BuOH (2 L × 3). The concentrated EtOAc fraction (16 g) was subjected to silica gel (SiO<sub>2</sub>) column (8 × 15 cm) chromatography (c.c.) and eluted with *n*-hexane–EtOAc (8:1→6:1→4:1, v/v) and CHCl<sub>3</sub>–MeOH (10:1→6:1→3:1→1:1, v/v). Each eluant was monitored by thin-layer chromatography (TLC), and 23 fractions (DBLE-1 to DBLE-23) were obtained. Fraction DBLE-9 (621.5 mg) was subjected to ODS c.c. and eluted with MeOH–H<sub>2</sub>O (5:1→12:1, v/v) to yield 23 subfractions (DBLE-9-1 to DBLE-9-23). Compound **1** [DBLE-9-21, 9.4 mg, *R<sub>f</sub>* 0.27 on the TLC (RP-18 F<sub>254</sub>s) in acetone–acetonitrile 1:2] and compound **2** [DBLE-9-22, 40.6 mg, *R<sub>f</sub>* 0.25 on the TLC (RP-18 F<sub>254</sub>s) in acetone–acetonitrile 1:2] were isolated. Fraction DBLE-21 (5.8 g) was subjected to SiO<sub>2</sub> c.c. and eluted with CHCl<sub>3</sub>–MeOH (20:1→10:1, v/v) to yield 18 subfractions (DBLE-21-1 to DBLE-21-18). Subfraction DBLE-21-13 (795.9 mg) was subjected to ODS c.c. and eluted with MeOH–H<sub>2</sub>O (1:1.5→6:1, v/v) to yield 23 subfractions (DBLE-21-13-1 to DBLE-21-13-23). Compound **3** [DBLE-21-13-22, 75 mg, *R<sub>f</sub>* 0.20 on the TLC (RP-18 F<sub>254</sub>s) in MeOH–H<sub>2</sub>O 25:1] was isolated. Subfraction DBLE-21-8 (852.7 mg) was subjected to ODS c.c. and eluted with acetone–H<sub>2</sub>O (1:1.5→1:1, v/v) to yield 18 subfractions (DBLE-21-8-1 to DBLE-21-8-18). Subfraction DBLE-21-8-16 (47.8 mg) underwent flash chromatography over silica gel (Biotage® SNAP cartridge KP-Sil – 10 g) and was eluted with *n*-hexane–EtOAc (4:1, v/v) to yield 14 subfractions (DBLE-21-8-16-1 to DBLE-21-8-16-14), which ultimately yielded compound **4** [DBLE-21-8-16-9, 6.7 mg, *R<sub>f</sub>* 0.68 on the TLC (SiO<sub>2</sub> F<sub>254</sub>) *n*-hexane–EtOAc 1:3] and compound **5** [DBLE-21-8-16-12, 9.2 mg, *R<sub>f</sub>* 0.60 on the TLC (SiO<sub>2</sub> F<sub>254</sub>) in *n*-hexane–EtOAc 1:3].

Compounds **1**, **2**, and **3** were identified as stigmasterol,  $\beta$ -sitosterol, and daucosterol, respectively, from the comparison of collected spectroscopic data with that of authentic samples and literature [7–9]. FAB-MS, <sup>1</sup>H NMR, <sup>13</sup>C NMR, and 2D NMR (gCOSY, gHSQC, and gHMBC) were used to identify compounds **4** and **5**.

**Stigmast-5-en-3 $\beta$ ,7 $\beta$ -diol (4)**, white powder, mp 157–158°C, [ $\alpha$ ]<sub>D</sub><sup>25</sup> –4° (c 0.25, CHCl<sub>3</sub>); positive FAB-MS: *m/z* 431 [M + H]<sup>+</sup>. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>,  $\delta$ , ppm, J/Hz): 5.26 (1H, br.s, H-6), 3.83 (1H, br.d, J = 7.6, H-7), 3.55 (1H, m, H-3), 1.02 (3H, s, H-19), 0.90 (3H, d, J = 6.4, H-21), 0.85 (3H, t, J = 6.6, H-29), 0.81 (3H, d, J = 7.6, H-26), 0.79 (3H, d, J = 7.2, H-27), 0.67 (3H, s, H-18). <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>,  $\delta$ , ppm): 143.37 (C-5), 125.54 (C-6), 73.36 (C-7), 71.44 (C-3), 55.97 (C-14), 55.39 (C-17), 48.29 (C-9), 45.88 (C-24), 42.96 (C-13), 41.75 (C-4), 40.95 (C-12), 39.60 (C-8), 36.98 (C-1), 36.48 (C-10), 36.15 (C-20), 31.98 (C-22), 31.67 (C-2), 29.75 (C-23), 28.61 (C-16), 26.45 (C-15), 26.16 (C-25), 22.76 (C-28), 21.15 (C-11), 19.89 (C-27), 19.23 (C-26), 19.09 (C-21), 18.9 (C-19), 12.06 (C-29), 11.91 (C-18).

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**Stigmast-5-en-3 $\beta$ ,7 $\alpha$ -diol (5)**, white powder, mp 219–220°C,  $[\alpha]_D^{25}$   $-45^\circ$  (*c* 0.25, CHCl<sub>3</sub>); positive FAB-MS: *m/z* 431 [M + H]<sup>+</sup>. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>,  $\delta$ , ppm, J/Hz): 5.58 (1H, d-like, J = 4.8, H-6), 3.82 (1H, br.s, H-7), 3.59 (1H, m, H-3), 0.97 (3H, s, H-19), 0.90 (3H, d, J = 6.4, H-21), 0.84 (3H, t, J = 6.8, H-29), 0.81 (3H, d, J = 8.0, H-26), 0.79 (3H, d, J = 8.0, H-27), 0.66 (3H, s, H-18). <sup>13</sup>C NMR spectrum (100 MHz, CDCl<sub>3</sub>,  $\delta$ , ppm): 146.13 (C-5), 123.79 (C-6), 71.33 (C-3), 65.36 (C-7), 49.48 (C-14), 55.73 (C-17), 45.85 (C-24), 42.29 (C-9), 42.18 (C-13), 42.04 (C-4), 39.21 (C-12), 37.56 (C-8), 37.44 (C-1), 37.05 (C-10), 36.15 (C-20), 33.74 (C-22), 31.42 (C-2), 29.76 (C-23), 28.19 (C-16), 25.95 (C-15), 24.32 (C-25), 23.02 (C-28), 20.74 (C-11), 19.85 (C-27), 19.03 (C-26), 18.84 (C-21), 18.30 (C-19), 12.06 (C-29), 11.71 (C-18).

Leafy culms of *D. bipinnata* contained stigmasterol (**1**),  $\beta$ -sitosterol (**2**), daucosterol (**3**), stigmast-5-en-3 $\beta$ ,7 $\beta$ -diol (**4**), and stigmast-5-en-3 $\beta$ ,7 $\alpha$ -diol (**5**). All of these sterols were reported for the first time from the plant of the monotypic genus *Desmostachya*.

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## REFERENCES

1. Z. Y. Wu, P. H. Raven, and D. Y. Hong, *Flora of China-Poaceae*, Science Press, Beijing, Vol. **22**, 2006, 458 pp.
2. IUCN-Nepal, *National Register of Medicinal and Aromatic Plants*, International Union for Conservation of Nature (IUCN), Kathmandu, 2004, 202 pp.
3. S. Panda, N. S. K. Choudhury, B. R. Behera, S. K. Mahapatra, and B. C. Behera, *J. Teaching Res. Chem. Gayatri College of Pharmacy*, Orissa, India, **15**, 47 (2008).
4. K. V. S. R. G. Prasad, D. Sujatha, and K. Bharathi, *Pharmacognosy Rev.*, Medknow Publication, India, **1**, 175 (2007).
5. A. S. Awaad, N. H. Mohamed, D. J. Maitland, and G. A. Soliman, *Rec. Nat. Prod.*, **2**, 76 (2008).
6. M. A. Ramadan and N. A. Safwat, *Aust. J. Basic Appl. Sci.*, **3**, 2270 (2009).
7. P. Forgo and K. E. Kover, *Steroids*, **69**, 43 (2004).
8. X. Zhang, P. Geoffroy, M. Miesch, J. D. David, F. Raul, D. Aoude-Werner, and E. Marchioni, *Steroids*, **70**, 886 (2005).
9. D. Y. Lee, S. J. Lee, H. Y. Kwak, L. Jung, J. Heo, S. Hong, G. W. Kim, and N. I. Baek, *J. Microbiol. Biotechnol.*, **19**, 1328 (2009).